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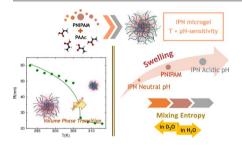
Swelling of responsive-microgels: experiments versus models



Valentina Nigro^{a,b,*}, Roberta Angelini^{a,b}, Monica Bertoldo^c, Barbara Ruzicka^{a,b}

- a Dipartimento di Fisica, Sapienza Università di Roma, P.le Aldo Moro 5, 00185 Roma, Italy
- b Istituto dei Sistemi Complessi del Consiglio Nazionale delle Ricerche (ISC-CNR), UOS Sapienza, Pz.le Aldo Moro 5, 00185 Roma, Italy
- ^c Istituto per i Processi Chimico-Fisici del Consiglio Nazionale delle Ricerche (IPCF-CNR), Area della Ricerca, Via G.Moruzzi 1, I-56124 Pisa, Italy

GRAPHICAL ABSTRACT



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ABSTRACT

Interpenetrated Polymer Network (IPN) microgels of PNIPAM and PAAc have been investigated and the experimental data have been compared with theoretical models from the Flory–Rehner theory. We confirm that the swelling behavior of PNIPAM microgels is well described by this theory by considering the second order approximation for the volume fraction ϕ dependence of the Flory parameter $\chi(\phi)$. Indeed the Volume-Phase Transition (VPT) of the PNIPAM-PAAc IPN microgel at neutral conditions and in D_2O solvents can be well described only considering a third-order approximation. Interestingly we empirically find that sharper is the transition higher is the order of the $\chi(\phi)$ relation which has to be considered. Moreover the VPT can be experimentally controlled by tuning the polymer/solvent interactions through pH and solvent allowing to directly modify the delicate balance between energetic and entropic contributions and to explore the swelling behavior in a wide range of environmental conditions. In particular we find that the most advantageous condition for swelling is in water at acidic pH.

1. Introduction

Responsive microgels are aqueous dispersions of nanometre- or micrometre-sized hydrogel particles [1–3], able to swell and retain large amount of water in response to slight changes in the environmental conditions. The high responsiveness of this interesting class of smart materials makes responsive microgels attractive for several technological applications [4–9] and very good model systems for understanding the complex behaviors emerging in soft colloids [10–12].

Indeed they allow to modulate the interparticle potential and their effective volume fraction through easily accessible control parameters such as temperature, pH or solvent, leading to novel phase-behaviors [13,3,14,2,15], drastically different from those of conventional hard colloidal systems [16–23].

The most known responsive microgels are based on poly(N-iso-propylacrylamide) (PNIPAM), which is a thermo-sensitive polymer [24] with a Lower Critical Solution Temperature (LCST) in water at about 306 K. At room temperature indeed, the polymer is hydrophilic and

^{*} Corresponding author at: Dipartimento di Fisica, Sapienza Università di Roma, P.le Aldo Moro 5, 00185 Roma, Italy. E-mail address: valentina.nigro@uniroma1.it (V. Nigro).

strongly hydrated in solution, while it becomes hydrophobic above 306 K, where a coil-to-globule transition takes place. This gives rise to a Volume-Phase Transition (VPT) from a swollen to a shrunken state of any PNIPAM-based microgel [25]. This typical swelling/shrinking behavior has been shown to be the driving mechanism of the phase behavior of aqueous suspensions of PNIPAM microgels, since it affects the interactions between particles and provides good tunability of both softness and volume fraction as a function of temperature [26,2,1,3,13]. It has been recently shown that the microgel swelling/shrinking behavior can be strongly affected by concentration [27,13], solvents [28] and internal structure and composition (such as number and distribution of cross-linking points [29,30] and core–shell structure [31,32]) or by introducing additives into the PNIPAM network [31].

In this context PNIPAM microgels containing another specie as comonomer or interpenetrated polymer are even more interesting, as a more complex scenario comes out. In particular, addition of poly(acrilic acid) (PAAc) to PNIPAM microgel provides an additional pH-sensitivity to the thermo-responsive microgel. Therefore their temperature dependent volume phase transition can be controlled by the content of AAc monomer [33,34], by the pH [35–40] or the ionic strength [35,41] of the suspension, since it strongly depends on the effective charge density.

In this framework the synthesis procedure plays a crucial role, since the response of PNIPAM/PAAc microgels is strictly related to the mutual interference between the two monomers [35,36,38,41,32,42-44]. In particular interpenetration of hydrophilic PAAc into the PNIPAM microgel network (IPN PNIPAM-PAAc microgel) [33,37,45-48], has little influence on the coil-to-globule transition temperature of the PNIPAM chains [34], leaving the temperature dependence of the VPT almost unchanged with respect to the case of pure PNIPAM microgel. Nevertheless the different solubility of PAAc at acidic and neutral pH, introduces an additional parameter to control the delicate balance between polymer/polymer and polymer/solvent interactions. At acidic pH the PAAc chains are not effectively solvated by water and H-bonds between the carboxylic (COOH-) groups of PAAc and the isopropyl (CONH-) groups of PNIPAM are favored [49,40]; at neutral pH, instead, the balance between PNIPAM/PAAc and water/PAAc H-bonds is reversed: both compounds are therefore well solvated and water mediates their interaction, making the two networks completely independent one to each other.

The typical swelling behavior of these thermo-responsive microgels has attracted great interest in the last years, with an increasing number of theoretical works aimed at describing this phenomenon [50–54]. In particular, for PNIPAM-based microgels it is well known that the swelling behavior can be well described by the classic Flory–Rehner theory [50], which has been extended to take into account the cross-linker influence on the polymer-solvent interaction [55–57]. It has been shown that the driving force for swelling can be estimated from the properties of linear PNIPAM solutions, while the microgel elasticity opposing swelling is mainly due to the network topology dependent on the cross-linker concentration. On the other hand, it has been shown that the swelling behavior is highly influenced by the effective charge density, which in PNIPAM-PAAc IPN microgels can be experimentally controlled by the content of AAc monomer [33,34] or by the pH of the suspension

In this framework the swelling behavior of aqueous suspensions of PNIPAM-PAAc IPN microgels as a function of temperature, pH and concentration has been extensively investigated by our group through Dynamic Light Scattering (DLS) and Small-Angle Neutron Scattering (SANS) [39,40]. In particular it has been observed that, in the high dilution regime (where the interparticle interactions are negligible and phase separation does not occur), a VPT around 305–307 K occurs. However interesting differences depending on pH and H/D isotopic substitution have been found, suggesting that H-bonding plays a nontrivial role. Nevertheless there is a weak understanding of the real fundamental reasons of their peculiar behaviors. Therefore the aim of

this work is to compare our experimental data with theoretical models accounting for the sharpness of the transition, in order to describe the swelling behavior of PNIPAM-PAAc IPN microgel, where the VPT can be experimentally controlled by tuning pH or acrylic acid content and by isotope substitution in the solvent.

2. Experimental methods

2.1. Sample preparation

2.1.1. Materials

N-isopropylacrylamide (NIPAM) from Sigma–Aldrich and N,N'-methylene-bis-acrylamide (BIS) from Eastman Kodak were recrystallized from hexane and methanol, respectively. Acrylic acid (AAc) from Sigma–Aldrich was purified by distillation (40 mmHg, 337 K). Sodium dodecyl sulphate (SDS), 98% purity, potassium persulfate (KPS), 98% purity, ammonium persulfate, 98% purity, N,N,N',N'-tetramethylethylenediamine (TEMED), 99 % purity, ethylenediaminetetraacetic acid (EDTA), NaHCO3, were all purchased from Sigma–Aldrich and used as received. Ultrapure water (resistivity: 18.2 $M\Omega$ /cm at 298 K) was obtained with Sariumlapro Ultrapure Water purification Systems, Sartorius Stedim from demineralized water. D_2O (99.9 atom %) from Sigma–Aldrich. All other solvents were RP grade (Carlo Erba) and were used as received. A dialysis tubing cellulose membrane, HCWO 14,000 Da, from Sigma–Aldrich, was cleaned before use by washing in distilled water and NaHCO3/EDTA solution.

2.1.2. Synthesis of PNIPAM and IPN microgels

The IPN microgel was synthesized by a sequential free radical polymerization method [33]. In the first step PNIPAM micro-particles synthesized with $(4.0850 \pm 0.0001) \,\mathrm{g}$ of NIPAM. (0.0695 ± 0.0001) g of BIS and (0.5990 ± 0.0001) g of SDS in 300 mL of ultrapure water. The solution was purged with nitrogen for 30 min, heated at (343.0 \pm 0.3) K and the polymerization was initiated with (0.1780 ± 0.0001) g of KPS. After 4 h the resulting PNIPAM microgel was purified by dialysis against distilled water for 2 weeks. For the synthesis of the second network, (65.45 \pm 0.01) g of the recovered PNIPAM dispersion (1.02 wt% and 80 nm diamter at 298 K) was diluted with ultrapure water up to a volume of 320 mL. (0.50 ± 0.01) g of BIS were mixed and the mixture was purged with nitrogen for 1 h. 2.3 mL of AAc and (0.2016 ± 0.0001) g of TEMED and (0.2078 ± 0.0001) g of ammonium persulfate were added and the reaction was carried out for 65 min at (295 \pm 1) K. The obtained IPN microgel was purified by dialysis against distilled water for 2 weeks, and then lyophilized to constant weight. The PAAc second network amount in the IPN was 31.5% by gravimetric analysis and the acrylic acid monomer units content 22% by acid/base titration. The remaining 9.5% being ascribed to BIS. The sample was lyophilized and redispersed again in H₂O or in D₂O by magnetic stirring for 1 day to obtain the final suspension at the required weight concentration. Samples at different weight concentrations (wt.%), in the following referred to simply as C_w , were obtained by dilution and by keeping constant the molar ratio in H₂O and D₂O. Therefore in the following we will refer to the weight concentration in H2O.

2.2. DLS set-up and data analysis

DLS measurements have been performed with a multiangle light scattering setup. The monochromatic and polarized beam emitted from a solid state laser (100 mW at $\lambda=642$ nm) is focused on the sample placed in a cylindrical VAT for index matching and temperature control. The scattered intensity is simultaneously collected by single mode optical fibers at five different scattering angles, namely $\theta=30^\circ, 50^\circ, 70^\circ, 90^\circ, 110^\circ,$ corresponding to different scattering vectors Q, according to the relation $Q=(4\pi n/\lambda)\sin(\theta/2)$. In this way one can simultaneously measure the normalized intensity autocorrelation

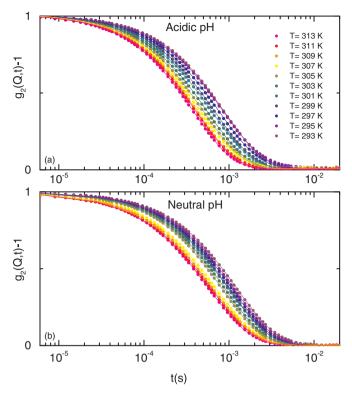


Fig. 1. Normalized intensity autocorrelation functions of an IPN sample at $C_w = 0.10\%$, at (a) acidic and (b) neutral pH and $\theta = 90^\circ$ for the indicated temperatures. Full lines are fits according to Eq. (1).

function $g_2(Q, t) = \langle I(Q, t)I(Q, 0) \rangle / \langle I(Q, 0) \rangle^2$ at five different Q values with a high coherence factor close to the ideal unit value. Measurements have been performed as a function of temperature in the range $T = (293 \div 313)$ K across the VPT at low weight concentrations, at both acidic and neutral pH. Reproducibility has been tested by repeating measurements several times.

In Fig. 1 the typical behavior of the normalized intensity auto-correlation functions for an IPN sample at weight concentration C_w = 0.10%, at acidic (\approx 5) and neutral (\approx 7) pH and θ = 90° for the indicated temperatures is shown.

As commonly known, the intensity correlation function of most colloidal systems is well described by the Kohlrausch–Williams–Watts expression [58,59]:

$$g_2(Q, t) = 1 + b[(e^{-t/\tau})^{\beta}]^2$$
 (1)

where b is the coherence factor, τ is an "effective" relaxation time and β describes the deviation from the simple exponential decay ($\beta=1$) usually found in monodisperse systems and gives a measure of the distribution of relaxation times. Many glassy materials show a stretching of the correlation functions (here referred to as "stretched behavior") characterized by an exponent $\beta<1$. In the case of Brownian diffusion it is possible to relate the relaxation time to the translational diffusion coefficient D_t through the relation $\tau=1/(Q^2D_t)$. In the limit of non interacting spherical particles the hydrodynamic radius is given by the Stokes Einstein relation

$$R = K_B T / 6\pi \eta D_t \tag{2}$$

where η is the sample viscosity, T is the sample temperature and k_B is the Boltzmann constant. Indeed, once the viscosity η is known, this relation allows to calculate the hydrodynamic radius from the diffusion coefficient. In particular the hydrodynamic radii of this work have been calculated by using the viscosity of the solvent, since samples are in the high dilution limit.

3. Swelling theory

Responsive microgels based on thermo-sensitive polymers, such as those investigated in this work, exhibit a complex and exotic behavior driven by the response to temperature changes. It is well known that the VPT of microgel particles is closely related to the coil-to-globule transition of the polymer chains composing the microgel network and it has been extensively explained by the Flory–Rehner theory [50]. At equilibrium the net osmotic pressure in a microgel, consisting of a mixing π_{mixing} and an elastic $\pi_{elasticity}$ contribution, is equal to zero

$$\pi = \pi_{\text{mixing}} + \pi_{\text{elasticity}} = 0 \tag{3}$$

This implies that [50]

$$\ln(1 - \phi) + \phi + \chi \phi^2 + \frac{\phi_0}{N} \left[\left(\frac{\phi}{\phi_0} \right)^{1/3} - \frac{1}{2} \frac{\phi}{\phi_0} \right] = 0$$
 (4)

where χ is the Flory polymer–solvent energy parameter (empirically given as a function of temperature and microgel weight fraction), N is the average number of segments between two neighboring cross-linking points in the microgel network, ϕ is the polymer volume fraction within the particle and ϕ_0 is the polymer volume fraction in the reference state, where the conformation of the network chains is the closest to that of unperturbed Gaussian chains. Typically this reference state is taken as the shrunken one and ϕ_0 is the polymer volume fraction corresponding to the shrunken state. For isotropic swelling, ϕ can be related to the particle size R as

$$\frac{\phi}{\phi_0} = \left(\frac{R_0}{R}\right)^3 \tag{5}$$

where R_0 is the particle diameter at the reference state (i.e. the shrunken state) and R is the particle diameter at a given state.

Therefore from Flory–Rehner theory we obtain the relation between the particle size and the other relevant parameter, thus giving details about the thermodynamics of the swelling/deswelling mechanism. In this framework the main role is played by the Flory χ parameter, reflecting the influence of the energy and entropy changes during the mixing process. It is indeed defined as the free energy change per solvent molecule when both polymer–polymer and solvent–solvent contacts are replaced by polymer–solvent contacts [50,54,60]

$$\chi = \frac{\Delta G}{k_B T} = \frac{\Delta H - T \Delta S}{k_B T} = \frac{1}{2} - A \left(1 - \frac{\theta}{T} \right) \tag{6}$$

where k_B is the Boltzmann constant, T is the temperature and A and θ are defined as

$$A = \frac{2\Delta S + k_B}{2k_B} \tag{7}$$

$$\theta = \frac{2\Delta H}{2\Delta S + k_R} \tag{8}$$

A rich behavior is observed depending on χ , which embodies the temperature dependence of the swelling behavior. For $T=\theta$, the Flory parameter is $\chi=1/2$, corresponding to the equilibrium conditions: polymer coils act like ideal chains and the excess of osmotic pressure of mixing between polymer and solvent is zero. For values $\chi<1/2$, the solvent is a good solvent for the polymer, interactions between polymer segments and solvent molecules are energetically favorable and allow polymer coils to expand. In these conditions the mixing contribution to the osmotic pressure is positive, thus promoting swelling. For $\chi>1/2$ the solvent is a poor solvent, hence polymer–polymer self-interactions are preferred and the polymer coils contract, thus promoting deswelling.

However the original Flory–Rehner theory is not able to describe the discontinuous transition observed in soft colloidal microgels due to the high complexity of their structure. It has been shown [61,54] that

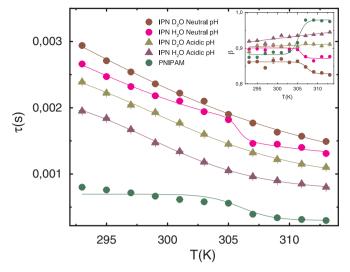


Fig. 2. Relaxation time and stretching parameter (inset) as a function of temperature for H_2O and D_2O suspensions of IPN microgels at acidic and neutral pH, compared with aqueous suspensions of PNIPAM microgels at the same weight concentration ($C_w = 0.10\%$). Full lines are guides for eyes.

exchange interactions must be described by accounting for interactions of higher order than contacts between molecules. This means that the χ parameter as a function of the volume fraction ϕ , has to be written as a power series expansion

$$\chi = \chi_1(T) + \chi_2 \phi + \chi_3 \phi^2 + \chi_4 \phi^3 + \cdots$$
 (9)

where χ_1 is the zero-order original Flory parameter of Eq. (6) and χ_2 , χ_3 , χ_4 , ... are temperature independent coefficients which introduce additional terms in the equation of state (Eq. (4)).

This empirical model has been extensively used for describing the swelling behavior of homopolymeric PNIPAM microgels [60,54]. However we propose to apply the same theory to non-homogeneous systems, such as PNIPAM-PAAc IPN microgel, by assuming that χ is an effective mean parameter accounting for polymer/polymer interactions within each network, polymer/polymer interactions between different networks and polymer/solvent interactions.

4. Results and discussions

In Fig. 2 we report the comparison between the temperature dependence of the relaxation time, as obtained through a fit with Eq. (1), for $\rm D_2O$ and $\rm H_2O$ suspensions of IPN microgels, at both acidic and neutral pH and for aqueous suspension of PNIPAM microgels, at the same fixed concentration ($C_w = 0.10\%$). In all the investigated samples a dynamical transition associated to the VPT, from a swollen to a shrunken state, is evidenced [39]. The relaxation time slightly decreases as temperature increases, until the transition is approached around T = 305 K. Above this Volume-Phase Transition Temperature (VPTT) it decreases to its lowest values, corresponding to the shrunken state.

However interesting differences between pure PNIPAM microgels and PNIPAM-PAAc IPN microgels can be noticed. In particular, the VPT of IPN microgels is strongly affected by the pH of the solution, due to the different solubility of PAAc at acidic and neutral pH. A first evidence of this pH-sensitivity is observed in the temperature behavior of the normalized intensity autocorrelation functions (Fig. 1), since an almost continuous transition at acidic pH (Fig. 1(a)) and a sharp transition at neutral pH (Fig. 1(b)) are observed around 305–307 K. Indeed as temperature increases, the relaxation time slowly decreases up to the VPTT, while above this temperature different behaviors come out depending on pH and solvent. For $\rm H_2O$ samples at acidic pH an almost continuous transition is observed. At neutral pH, instead, the H-bonds

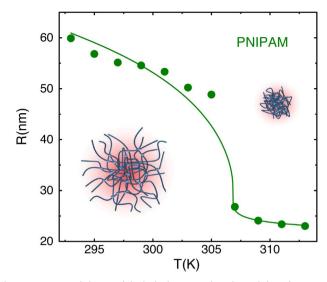


Fig. 3. Temperature behavior of the hydrodynamic radius obtained through DLS measurements for aqueous suspensions of PNIPAM microgels at $C_w = 0.10\%$. Solid line is the fit obtained through Eq. (10).

interactions between the carboxylic groups of PAAc and PNIPAM are reduced and a sharp transition to lower values of τ is observed. Moreover, although the main features of the dynamical behavior for H2O suspensions of PNIPAM-PAAc IPN microgels [39] are preserved under isotopic substitution, some differences can be observed in D₂O samples: the sharpness of the transition is widely reduced in D2O with respect to H₂O suspensions at neutral pH and the relaxation times are always higher in D₂O than in H₂O, suggesting a slowing down of the dynamics under isotopic substitution, probably due to the higher viscosity of D₂O compared to H_2O . Moreover the stretching parameter β (inset of Fig. 2) provides useful details on samples polydispersity. Indeed for PNIPAM microgels $\beta \approx 0.9$ indicates slightly stretched correlation curves and an almost monodisperse sample. For IPN microgels the scenario is more complex: at acidic pH $\beta \approx 0.9$ and the sample is mainly monodisperse in all the investigated temperature range, while at neutral pH it decreases with temperature up to $\beta \approx 0.8$, indicating more polydisperse samples above the VPTT.

All these features can be well visualized by looking at the temperature behavior of the hydrodynamic radius, as obtained from DLS measurements in the high dilution limit, where interparticle interactions can be neglected and the Stokes-Einstein relation (Eq. (2)) can be used. In Fig. 3 the temperature behavior of the hydrodynamic radius for aqueous suspensions of PNIPAM microgels at weight concentration $C_w = 0.10\%$ is shown. A cross-over from a swollen to a shrunken state is observed at $T \approx 306$ K. It is well known that the swelling behavior of PNIPAM microgels can be described by the Flory-Rehner theory, which well characterizes a discontinuous transition by assuming the volume fraction dependence of the Flory χ parameter (Eq. (6)). According to previous studies for PNIPAM microgels [54], a good agreement between theory and experiments is obtained with a second-order approximation of the $\chi(\phi)$ relation (Eq. (6)). Therefore by inverting Eq. (4) it is possible to explicit the temperature dependence on the particle radius

$$T = \frac{-A\phi_0^2 \left(\frac{R_0}{R}\right)^6 \theta}{\ln\left[1 - \phi_0 \left(\frac{R_0}{R}\right)^3\right] + \phi_0 \left(\frac{R_0}{R}\right)^3 + \phi_0^2 \left(\frac{R_0}{R}\right)^6 \left(\frac{1}{2} - A\right) + \chi_2 \phi_0^3 \left(\frac{R_0}{R}\right)^9} + \chi_3 \phi_0^4 \left(\frac{R_0}{R}\right)^{12} + \frac{\phi_0}{N} \left[\frac{R_0}{R} - \frac{1}{2} \left(\frac{R_0}{R}\right)^3\right]}$$
(10)

The solid line in Fig. 3 is the best fit of our experimental data through Eq. (10), with $\phi_0 \approx 0.8$, returning realistic fitting parameters.

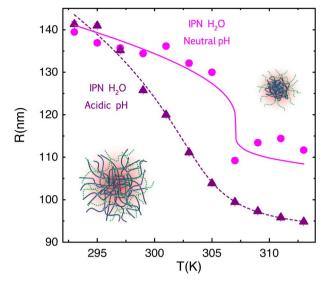


Fig. 4. Temperature behavior of the hydrodynamic radius obtained through DLS measurements for $\rm H_2O$ suspensions of IPN microgels at $C_w=0.10\%$. Dashed line is the fit obtained through Eq. (10) with the second-order approximation of the χ parameter and solid line is the fit obtained through Eq. (11) with the third-order approximation of the χ parameter.

In particular the value of the Flory temperature $\theta = (306.7 \pm 0.3)$ K is close to the VPTT, thus confirming that the volume phase transition is strictly related to the transition from good to poor solvent. Furthermore we find negative values of A, leading to negative values of ΔS and ΔH , as expected for systems with a LCST and confirming that for PNIPAM microgels the transition in the solvent quality is driven by order-disorder processes due to hydrophobic interactions [54].

As the acrylic acid is introduced within the PNIPAM network, the swelling capability is reduced with respect to PNIPAM microgels and an additional control parameter is introduced. In Fig. 4 the temperature behavior of the hydrodynamic radius for aqueous suspensions of IPN microgels at acidic and neutral pH is reported. The presence of PAAc in IPN microgels allows to modulate the sharpness and the amplitude of the VPT by changing pH or, equivalently, by deprotonating the sample. In particular at neutral pH the transition is sharper and narrower with respect to acidic pH, where a smoother and wider transition is observed, indicating a more hydrophobic system [39]. This behavior can be interpreted in terms of the role played by the PAAc in IPN microgels [40]. At low pH the acrylic acid is insoluble in water and is mainly in protonated state, therefore H-bonds between its carboxylic (COOH-) groups and the amidic (CONH-) groups of PNIPAM are formed and IPN microgels are strongly hydrophobic. Therefore when heated above the VPTT, due to the increased hydrophobic interactions, IPN microgels expel a large amount of water giving rise to a very dense shrunken state. At neutral pH the carboxylic groups of PAAc are deprotonated, leading to a strong charge repulsion which limits the formation of H-bonds with PNIPAM so that the two networks result independent and the sharpness of the transition is partially restored with respect to the case of pure PNIPAM.

Solid and dashed lines in Fig. 4 are the best fits of our experimental data, with $\phi_0 \approx 0.8$. In particular Eq. (10) well describes our data only at acidic pH, where the transition is smooth and the ϕ dependence of the Flory χ parameter is well approximated by the second-order approximation of Eq. (9). In this case the value of the Flory temperature $\theta = (304.8 \pm 0.2)$ K is close to the VPTT for IPN microgels at acidic pH [39], which is expected to be slightly shifted backward with respect to pure PNIPAM microgels. Furthermore we find negative values of A, indicating that the fundamental mechanism of the mixing process is not affected as the acrylic acid is introduced. These results confirm that at the IPN microgel structure can be modeled as an effective average network, indicating that the PNIPAM and PAAc networks are tightly

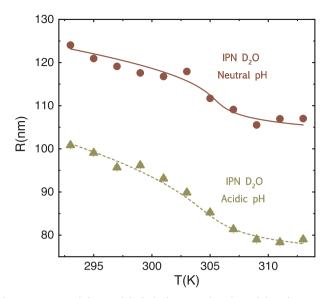


Fig. 5. Temperature behavior of the hydrodynamic radius obtained through DLS measurements, for D_2O suspensions of IPN microgels at $C_w=0.10\%$. Solid and dashed lines are the fit obtained through Eq. (11) with the third-order approximation of the χ parameter.

bound to each other.

At neutral pH instead the VPT becomes sharp and discontinuous and the description of the data through Eq. (10) fails, suggesting that a second-order approximation of the χ parameter is no more sufficient. We decided therefore to extend Eq. (10) considering the third-order expansion of the $\chi(\phi)$ relation (Eq. (9)):

$$T = \frac{-A\phi_0^2 \left(\frac{R_0}{R}\right)^6 \theta}{\ln\left[1 - \phi_0 \left(\frac{R_0}{R}\right)^3\right] + \phi_0 \left(\frac{R_0}{R}\right)^3 + \phi_0^2 \left(\frac{R_0}{R}\right)^6 \left(\frac{1}{2} - A\right) + \chi_2 \phi_0^3 \left(\frac{R_0}{R}\right)^9} + \chi_3 \phi_0^4 \left(\frac{R_0}{R}\right)^{12} + \chi_4 \phi_0^5 \left(\frac{R_0}{R}\right)^{15} + \frac{\phi_0}{N} \left[\frac{R_0}{R} - \frac{1}{2} \left(\frac{R_0}{R}\right)^3\right]}$$
(11)

By using this relation for fitting our experimental data, we find a good agreement between theory and experiments as shown in Fig. 4 at neutral pH. In particular we obtain values of the Flory temperature $\theta = (306.0 \pm 0.2)$ K close to the VPTT for neutral pH samples, which is expected to be shifted to higher temperature with respect to acidic pH [39,40]. Moreover the obtained values of A are negative and lower than those found at acidic pH, confirming that mixing is governed by the LCST and suggesting that the entropic change during the mixing process is less significant than at acidic pH.

In addition the swelling behavior is slightly affected by H/D isotopic substitution in the solvent, since H-bondings play a crucial role. The temperature dependence of the hydrodynamic radii of D₂O suspensions of IPN microgels at both acidic and neutral pH and at the same weight concentration ($C_w = 0.10\%$), is shown in Fig. 5. The presence of the acrylic acid reduces the swelling capability of the microgel particles also in deuterated suspensions [33,37,38]. Nevertheless for D₂O suspensions at acidic pH the transition appears slightly sharper and the range of variability of R reduced with respect to H₂O suspensions, while at neutral pH the transition is smoother and the range of variability of R unchanged. This leads to no significant changes at acidic or neutral pH, in contrast with H₂O suspensions where the swelling behavior is highly sensitive to pH. This suggests that the balance between polymer/ polymer and polymer/solvent interactions strictly depends on the solvent and that in D2O less evident changes with pH are expected in the solvent quality. In this case a good agreement between theory and experiments is obtained with a third-order approximation of the $\chi(\phi)$ relation at both acidic and neutral pH. Solid and dashed lines in Fig. 5 are the best fits of our experimental data through Eq. (11), with

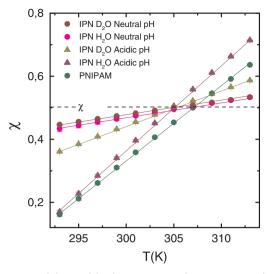


Fig. 6. Temperature behavior of the Flory χ_1 parameter for PNIPAM microgels and IPN microgels in H_2O and D_2O , at both acidic and neutral pH. Solid lines are guides to eyes.

Table 1 Values of the *A* parameter and Flory temperature θ as obtained from the best fit of the temperature behaviors of the hydrodynamic radii with Eq. (10) or Eq. (11) depending on the sample.

sample	A	θ (K)
PNIPAM IPN H_2O pH 5 IPN H_2O pH 7 IPN D_2O pH 5 IPN D_2O pH 5 IPN D_2O pH 7	-7.09 ± 0.06 -8.19 ± 0.06 -1.5 ± 0.2 -3.40 ± 0.03 -1.27 ± 0.07	306.7 ± 0.3 304.8 ± 0.2 306.2 ± 0.2 304.9 ± 0.4 305.0 ± 0.5

 $\phi_0 \approx 0.8$. We obtain values of the Flory temperature $\theta = (304.9 \pm 0.4)$ K at acidic pH and $\theta = (305.0 \pm 0.3)$ K at neutral pH, as expected from previous studies on these samples [39,40]. Moreover the obtained values of A are close to each other, thus confirming that in D₂O samples pH does not highly affects the swelling mechanism.

The differences in the swelling behaviors for the investigated samples can be well summarized by looking at the temperature behavior of the Flory χ_1 parameter. Indeed the χ_1 parameter reflects the influence of the energy and the entropy changes during the mixing process. It is

the zero-order approximation of Eq. (9) and is therefore given by Eq. (6), returning details about the delicate balance between energetic and entropic contributions. In Fig. 6 the behavior of χ_1 is reported for all the investigated samples. The obtained values of A and θ , reported in Table 1, are such that $0 < \chi_1 < 1$ as expected. Moreover we find $\chi_1 < 1/2$ below the VPTT and $\chi_1 > 1/2$ above the VPTT, thus indicating a transition from good to poor solvent across the volume-phase transition. However a different slope characterizes the χ_1 temperature behavior for different samples depending on pH and solvent, corresponding of the fitting parameters A and θ . In particular we find the largest slope for IPN microgels in water at acidic pH, indicating that mixing between polymer and solvent is favored and thus swelling promoted. This slope is even higher than for PNIPAM microgels, suggesting that the higher hydrophobicity due to the presence of PAAc may promote swelling. On the other hand the smallest slope of IPN microgels in water is found at neutral pH, meaning that the entropy changes are the least significant. Therefore the swelling process in H₂O is more favored at acidic than at neutral pH, being the system more hydrophobic. Moreover the χ_1 behavior also depend on solvent: in D₂O the changes between the χ_1 behaviors at acidic and neutral pH are less significant than in H₂O, confirming that the balance between polymer/ polymer and polymer/solvent interactions strictly depends on the solvent and therefore on the H-bondings.

From the temperature behavior of the hydrodynamic radius we experimentally calculate the χ parameter, which is plotted in Fig. 7 for H₂O samples as a function of T at acidic and neutral pH. At acidic pH (Fig. 7(a)) an almost continuous transition from the swollen to the shrunken state is observed and a second-order approximation of $\chi(\phi)$ well describes the microgel swelling. On the other hand at neutral pH the second-order approximation of the $\chi(\phi)$ relation (dashed lines in Fig. 7(b)) does not well reproduce the amplitude of the transition due to the increasing complexity of the microgel structure. Therefore a stronger ϕ dependence of γ is necessary to reproduce the sharp change across the VPT and a third-order approximation has to be taken into account (dotted line in Fig. 7(b)) to empirically capture the essential physics of the many-body interactions, whose exact nature is still lacking. However at low temperature the experimental points are well described with the theoretical solid line corresponding to the $\chi_1(T)$ behavior, as previously observed for PNIPAM microgels [54,60], even if the discrepancy between theoretical prediction and experimental points may suggest that particles are not fully swollen in the investigated temperature range. Therefore the Flory-Rehner theory well describes the main thermodynamic aspects of the volume-phase transition of

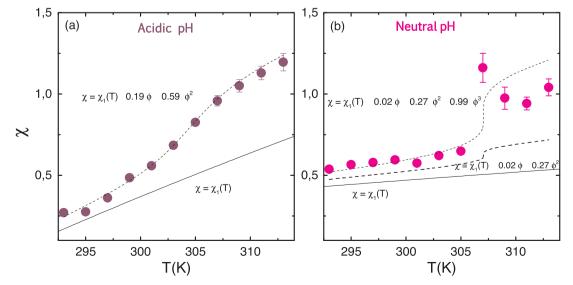


Fig. 7. Temperature behavior of the χ parameter for H₂O suspensions of IPN microgels at (a) acidic and (b) neutral pH. Solid lines correspond to the Flory χ_1 parameter, while dashed and dotted lines correspond to the theoretical predictions for $\chi(\phi)$.

PNIPAM-PAAc IPN microgels. However we empirically observe that a higher-order approximation of the $\chi(\phi)$ relation has to be considered as sharper is the transition. This is evident for IPN microgels in H_2O at neutral pH and for IPN microgels in D_2O at both acidic and neutral pH, where the discontinuous volume phase transition may be explained in terms of an increasing complexity of the microgel structure due to the breaking of the H-bonds between PNIPAM and PAAc networks. Therefore a third-order approximation of $\chi(\phi)$ is required to account for the balance between polymer/polymer and polymer/solvent interactions.

5. Conclusions

The swelling behavior of multi-responsive IPN microgels has been investigated and experimental data, obtained through DLS measurements, have been compared with models from the swelling theory. The discontinuous transitions can be described by introducing the dependence of the original Flory χ parameter from the volume fraction ϕ . In particular the swelling behavior of pure PNIPAM microgels is well described with a second-order approximation of the $\chi(\phi)$ relation, confirming previous results on these systems [54]. Also for PNIPAM-PAAc IPN microgels at acidic pH an almost continuous transition from a swollen to a shrunken state is observed and a good agreement between theory and experiments is obtained with a second-order approximation of the $\chi(\phi)$, reflecting the almost homogeneous structure of the microgel network due to the strong interactions between the PNIPAM and the PAAc networks. On the contrary at neutral pH the transition becomes sharp and discontinuous and a higher, third-order approximation of the $\gamma(\phi)$ relation has to be considered to account for the nonhomogeneity of the system when polymer/polymer interactions are replaced by polymer/solvent interactions. The same approximation is necessary to describe the swelling behavior of D₂O suspensions at both acidic and neutral pH. Indeed the VPT is affected by H/D isotopic substitution in the solvent, since the balance between polymer/polymer and polymer/solvent strictly depends on H-bondings. This leads to no significant changes with pH in D₂O suspensions and to an intermediate sharpness of the VPT with respect to H2O samples at both acidic and neutral pH. Indeed H-bond interactions are weaker in D2O than in H2O and at both acidic and neutral pH a higher degree of complexity has to be introduced. Details about the delicate balance between energetic and entropic contributions are embodied in the Flory χ_1 parameter. Its temperature behavior confirms that both for water and for heavy water the swelling is promoted at acidic pH, while at neutral pH the least advantageous condition for swelling is found. Therefore the observed behavior of $\chi_1(T)$ suggests that the swelling can be promoted or inhibited by controlling the polymer/solvent interactions through pH and solvent.

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References

- L.A. Lyon, A. Fernandez-Nieves, The polymer/colloid duality of microgel suspensions, Annu. Rev. Phys. Chem. 63 (2012) 25–43.
- [2] D. Paloli, P.S. Mohanty, J.J. Crassous, E. Zaccarelli, P. Schurtenberger, Fluid-solid transitions in soft-repulsive colloids, Soft Matter 9 (2013) 3000–3004.
- [3] P.S. Mohanty, D. Paloli, J.J. Crassous, E. Zaccarelli, P. Schurtenberger, Effective interactions between soft-repulsive colloids: experiments, theory and simulations, J. Chem. Phys. 140 (2014) 094901.
- [4] S.V. Vinogradov, Colloidal microgels in drug delivery applications, Curr. Pharm. Des. 12 (2006) 4703–4712.
- [5] M. Das, H. Zhang, E. Kumacheva, MICROGELS: old materials with new applications, Annu. Rev. Mater. Res. 36 (2006) 117–142.
- [6] J.S. Park, H.N. Yang, D.G. Woo, S.Y. Jeon, K.H. Park, Poly(N-isopropylacrylamide-co-acrylic acid) nanogels for tracing and delivering genes to human mesenchymal stem cells, Biomaterials 34 (2013) 8819–8834.

- [7] M. Hamidi, A. Azadi, P. Rafie, Hydrogel nanoparticles in drug delivery, Adv. Drug Deliv. Rev. 60 (2008) 1638–1649.
- [8] N.M.B. Smeets, T. Hoare, Designing responsive microgels for drug delivery applications, J. Polym. Sci. A Polym. Chem. 51 (2013) 3027–3043.
- [9] S. Su, Ali Md. Monsur, C.D.M. Filipe, Y. Li, R.H. Pelton, Microgel-based inks for paper-supported biosensing applications, Biomacromolecules 9 (2008) 935–9419.
- [10] C.N. Likos, N. Hoffmann, H. Löwen, A.A. Louis, Exotic fluids and crystals of soft polymeric colloids, J. Phys. Cond. Matter 14 (2002) 7681–7698.
- [11] P.E. Ramírez-González, M. Medina-Noyola, Glass transition in soft-sphere dispersions, J. Phys. Cond. Matter 21 (2009) 075101.
- [12] D.M. Heyes, S.M. Clarke, A.C. Brank, Interactions between microgel particles, Soft Matter 5 (2009) 2681–2685.
- [13] H. Wang, X. Wu, Z. Zhu, C.S. Liu, Z. Zhang, Revisit to phase diagram of poly(N-isopropylacrylamide) microgel suspensions by mechanical spectroscopy, J. Chem. Phys. 140 (2014) 024908.
- [14] T. Hellweg, C.D. Dewhurst, E. Brückner, K. Kratz, W. Eimer, Colloidal crystals made of poly(N-isopropylacrylamide) microgel particles, Colloid. Polym. Sci. 278 (2000) 972–978
- [15] J. Wu, B. Zhou, Z. Hu, Phase behavior of thermally responsive microgel colloids, Phys. Rev. Lett. 90 (4) (2003) 048304.
- [16] P.N. Pusey, W. van Megen, Phase behaviour of concentrated suspensions of nearly hard colloidal spheres, Nature 320 (1986) 340–342.
- [17] A. Imhof, J.K.G. Dhont, Experimental phase diagram of a binary colloidal hardsphere mixture with a large size ratio, Phys. Rev. Lett. 75 (1995) 1662–1665.
- [18] K.N. Pham, A.M. Puertas, J. Bergenholtz, S.U. Egelhaaf, A. Moussaïd, P.N. Pusey, A.B. Schofield, M.E. Cates, M. Fuchs, W.C.K. Poon, Multiple glassy states in a simple model system, Science 296 (2002) 104–106.
- [19] T. Eckert, E. Bartsch, Re-entrant glass transition in a colloid–polymer mixture with depletion attractions, Phys. Rev. Lett. 89 (2002) 125701.
- [20] P.J. Lu, E. Zaccarelli, F. Ciulla, A.B. Schofield, F. Sciortino, D.A. Weitz, Gelation of particle with short range attraction, Nature 453 (2008) 499–503.
- [21] C.P. Royall, S.R. Williams, T. Ohtsuka, H. Tanaka, Direct observation of a local structural mechanism for dynamical arrest, Nat. Mater. 7 (2008) 556–561.
- [22] B. Ruzicka, E. Zaccarelli, L. Zulian, R. Angelini, M. Sztucki, A. Moussaïd, T. Narayanan, F. Sciortino, Observation of empty liquids and equilibrium gels in a colloidal clav. Nat. Mater. 10 (2011) 56–60.
- [23] R. Angelini, E. Zaccarelli, F.A. de Melo Marques, M. Sztucki, A. Fluerasu, G. Ruocco, B. Ruzicka, Glass-glass transition during aging of a colloidal clay, Nat. Commun. 5 (2014) 4049.
- [24] R.H. Pelton, P. Chibante, Preparation of aqueous lattices with N-isopropylacrylamide. Colloids Surf. 20 (1986) 247–256.
- [25] J. Wu, G. Huang, Z. Hu, Interparticle potential and the phase behavior of temperature-sensitive microgel dispersions, Macromolecules 36 (2003) 440–448.
- [26] J. Mattsson, H.M. Wyss, A. Fernandez-Nieves, K. Miyazaki, Z. Hu, D. Reichman, D.A. Weitz, Soft colloids make strong glasses, Nature 462 (5) (2009) 83–86.
- [27] B.H. Tan, R.H. Pelton, K.C. Tam, Microstructure and rheological properties of thermo-responsive poly(N-isopropylacrilamide) microgels, Polymers 51 (2010) 3238–3243.
- [28] P.W. Zhu, D.H. Napper, Light scattering studies of poly(N-isopropylacrylamide) microgel particle in mixed water-acetic acid solvents, Macromol. Chem. Phys. 200 (1999) 1950-1955.
- [29] K. Kratz, W. Eimer, Swelling properties of colloidal poly(N-isopropylacrylamide) microgels in solution, Ber. Bunsenges. Phys. Chem. 102 (1998) 848–854.
- [30] K. Kratz, T. Hellweg, W. Eimer, Structural changes in PNIPAM microgel particles as seen by SANS, DLS and EM techniques, Polymer 42 (2001) 6631–6639.
- [31] T. Hellweg, C.D. Dewhurst, W. Eimer, K. Kratz, PNIPAM-co-polystyrene core-shell microgels: structure, swelling behavior, and crystallization, Langmuir 20 (2004) 4333–4335
- [32] Z. Meng, J.K. Cho, S. Debord, V. Breedveld, L.A. Lyon, Crystallization behavior of soft, attractive microgels, J. Phys. Chem. B 111 (2007) 6992–6997.
- [33] Z. Hu, X. Xia, Hydrogel nanoparticle dispersions with inverse thermoreversible gelation, Adv. Mater. 16 (4) (2004) 305–309.
- [34] J. Ma, B. Fan, B. Liang, J. Xu, Synthesis and characterization of poly(N-iso-propylacrylamide)/poly(acrylic acid) semi-IPN nanocomposite microgels, J. Colloid Interface Sci. 341 (2010) 88–93.
- [35] K. Kratz, T. Hellweg, W. Eimer, Influence of charge density on the swelling of colloidal poly(N-isopropylacrylamide-co-acrylic acid) microgels, Colloids Surf. A 170 (2000) 137–149.
- [36] K. Kratz, T. Hellweg, W. Eimer, Effect of connectivity and charge density on the swelling and local structure and dynamic properties of colloidal PNIPAM microgels, Ber. Bunsenges. Phys. Chem. 102 (11) (1998) 1603–1608.
- [37] X. Xia, Z. Hu, Synthesis and light scattering study of microgels with interpenetrating polymer networks, Langmuir 20 (2004) 2094–2098.
- [38] C.D. Jones, L.A. Lyon, Synthesis and characterization of multiresponsive core-shell microgels, Macromolecules 33 (2000) 8301–8303.
- [39] V. Nigro, R. Angelini, M. Bertoldo, V. Castelvetro, G. Ruocco, B. Ruzicka, Dynamic light scattering study of temperature and pH sensitive colloidal microgels, J. Non-Cryst. Solids 407 (2015) 361–366.
- [40] V. Nigro, R. Angelini, M. Bertoldo, F. Bruni, M.A. Ricci, B. Ruzicka, Local structure of temperature and ph-sensitive colloidal microgels, J. Chem. Phys. 143 (2015) 114004
- [41] W. Xiong, X. Gao, Y. Zao, H. Xu, X. Yang, The dual temperature/pH-sensitive multiphase behavior of poly(Nisopropylacrylamide-co-acrylic acid) microgels for potential application in in situ gelling system, Colloids Surf. B: Biointerfaces 84 (2011) 103–110.
- [42] L.A. Lyon, J.D. Debord, S.B. Debord, C.D. Jones, J.G. McGrath, M.J. Serpe, Microgel

- Colloidal Crystals, J. Phys. Chem. B 108 (2004) 19099-19108.
- [43] P. Holmqvist, P.S. Mohanty, G. Nägele, P. Schurtenberger, M. Heinen, Structure and dynamics of loosely cross-linked ionic microgel dispersions in the fluid regime, Phys. Rev. Lett. 109 (2012) 048302.
- [44] S.B. Debord, L.A. Lyon, Influence of particle volume fraction on packing in responsive hydrogel colloidal crystals, J. Phys. Chem. B 107 (2003) 2927–2932.
- [45] X. Xia, Z. Hua, M. Marquez, Physically bonded nanoparticle networks: a novel drug delivery system, J. Control. Release 103 (2005) 21–30.
- [46] J. Zhou, G. Wang, L. Zou, L. Tang, M. Marquez, Z. Hu, Viscoelastic behavior and in vivo release study of microgel dispersions with inverse thermoreversible gelation, Biomacromolecules 9 (2008) 142–148.
- [47] Z. Xing, C. Wang, J. Yan, L. Zhang, L. Li, L. Zha, pH/temperature dual stimuliresponsive microcapsules with interpenetrating polymer network structure, Colloid Polym. Sci. 288 (2010) 1723–1729.
- [48] X. Liu, H. Guo, L. Zha, Study of pH/temperature dual stimuli-responsive nanogels with interpenetrating polymer network structure, Polymers 61 (7) (2012) 1144–1150
- [49] E. Siband, Y. Tran, D. Hourdet, Thermoresponsive interpolyelectrolyte complexation: application to macromolecular assemblies, Macromolecules 44 (2011) 8185–8194
- [50] P.J. Flory, Principles of Polymer Chemistry, Cornell University, Ithaca, New York, 1953.
- [51] A.K. Lele, M.M. Hirve, M.V. Badiger, R.A. Mashelkar, Predictions of bound water content in poly (N-isopropylacrylamide) gel, Macromolecules 30 (1998) 157–159.
- [52] T. Hino, J.M. Prausnitz, Swelling equilibria for heterogeneous polyacrylamide gels,

- J. Appl. Polym. Sci. 62 (1996) 1635-1640.
- [53] K. Otake, H. Inomata, M. Konno, S. Saito, Thermal-analysis of the volume phasetransition with N-Isopropylacrylamide gels, Macromolecules 23 (1990) 283–289.
- [54] T. Lòpez-Leòn, A. Fernandez-Nieves, Macroscopically probing the entropic influence of ions: deswelling neutral microgels with salt, Phys. Rev. E 75 (2007) 1–7.
- [55] J.J. Crassous, A. Wittemann, M. Siebenbürger, M. Schrinner, M. Drechsler, M. Ballauff, Direct imaging of temperature-sensitive core-shell latexes by cryogenic transmission electron microscopy, Colloid Polym. Sci. 286 (2008) 805–812.
- [56] Y. Hertle, M. Zeiser, c. Hasenöhrl, P. Bush, T. Hellweg, Responsive P(NIPAM-co-NtBAM) microgels: Flory–Rehner description of the swelling behaviour, Colloid Polym. Sci. 288 (2010) 1047–1059.
- [57] A. Balaceanu, Y. Verkh, D.E. Demco, M. Möller, A. Pich, Correlated morphological changes in the volume temperature transition of core–shell microgels, Macromolecules 46 (2013) 4882–4891.
- [58] R. Kohlrausch, Thermoresponsive poly-(N-isopropylmethacrylamide) microgels: tailoring particle size by interfacial tension control, Pogg. Ann. Phys. Chem. 91 (1854) 179–214.
- [59] G. Williams, D.C. Watts, Non-symmetrical dielectric relaxation behavior arising from a simple empirical decay function, J. Chem. Soc. Faraday Trans. 66 (1970) 80–85
- [60] M. Shibayama, Y. Shirotami, H. Hirose, S. Nomura, Simple scaling rules on swollen and shrunken polymer gels, Macromolecules 30 (1997) 7307–7312.
- [61] B. Erman, P.J. Flory, Critical phenomena and transitions in swollen polymer networks and in linear macromolecules, Macromolecules 19 (1986) 2342–2353.